

# Immunohistochemical and Electron Microscope Analysis of Adenomas of the Thyroid Gland

## II. Adenomas With Specific Cytological Differentiation

W. Böcker, H. Dralle, G. Koch, K. de Heer, and J. Hagemann

Institute of Pathology (Director: Prof. Dr. G. Seifert), Surgical Department  
(Director: Prof. Dr. H.-W. Schreiber) and Radiological Department,  
Section of Nuclear Medicine (Chief: Prof. Dr. C. Schneider), University of Hamburg

**Summary.** Histological, immunohistochemical and electronmicroscopic studies of 12 human, scintigraphically “cold”, thyroid adenomas with specific cytological differentiation identified four different cell types: oxyphil cell, clear cell, ergastoplasm-rich cell and mitochondrion-rich cell.

The oxyphil tumor cell can be recognized light-microscopically by its large size and its eosinophilic granular cytoplasm. Most of these cells do not produce thyroglobulin. The ultrastructural characteristics of oxyphil cells are principally mitochondria in great numbers and many large lysosomes. Clear cell adenomas show a trabecular growth pattern. The tumor cells have an abundance of cytoplasm which contains small acidophilic granules. Immunohistochemically we were able to demonstrate thyroglobulin in small amounts within cytoplasmic granules and more extensively within the follicle lumina. Electronmicroscopically we observed a large number of smooth surfaced vacuoles of varying size, extraordinary large lysosomes and occasional cisternae of rough endoplasmic reticulum, the latter probably corresponding to the immune-histochemically identified thyroglobulin granules. The ergastoplasm-rich-cell adenomas, which to the best of our knowledge have not been previously described, show a predominantly micro-to normo-follicular architecture histologically without intrafollicular colloid. The cytoplasm of the ergastoplasm-rich cells reveals a strong positive thyroglobulin-staining reaction. The fine structure of these cells is characterized by the abundance of cisternae of the rough endoplasmic reticulum. The mitochondrion-rich-cell adenomas exhibited a microfollicular structure with an intensive acidophilic granular staining at the basal part of the tumor cells. Immunohistochemically and electronmicroscopically we found some morphologic and functional features which differentiate these cells from the oxyphil cell. Thyroglobulin was located predominantly in the apical portion of the cytoplasm in the mitochondrion-rich cells without sharp demarcation from the

luminar thyroglobulin. Electron microscopically fewer basal and laterally located mitochondria were seen in mitochondrion-rich cells compared with oxyphil cells. As we could not find any sign of functional activity in the oxyphilic, clear cell and ergastoplasm-rich cell adenomas we analysed those aspects of the lysosomal system not concerned with the enzymatic digestion of thyroglobulin.

**Key words:** Thyroid — Adenoma — Ultrastructure — Immunohistochemistry — Thyroglobulin — Lysosomes.

## Introduction

In a previous paper we examined cellular aspects of autonomous "hot" adenomas and scintigraphically "cold" adenomas (Dralle and Böcker, 1977). It was demonstrated that autonomous adenomas consisted of hypertrophic follicular cells while "cold" adenomas contain principle-(main-) cells that can not be distinguished morphologically from normal follicular cells. This publication presents histological, immunohistochemical and electronmicroscopic observations of 12 human thyroid adenomas with specific cytologic differentiation. In addition to the well known oxyphilic (Askanazy, 1898; Langhans, 1907; Wegelin, 1926; Lennox, 1948; Hamperl, 1950 and 1962; Toniatti et al., 1967; Klinck et al., 1970; Michel-Bechet et al., 1971; Kennedy and Thompson, 1974; Roediger, 1975) and clear cell adenoma (references see Stoll and Lietz, 1973), two further adenoma types presented: one type is composed of ergastoplasm-rich cells and the other of mitochondrion-rich cells.

The present study reveals new findings which might help to clarify some of the controversial questions regarding the cytogenesis and functional properties of these tumors. Lysosomes are directly related to the secretion of thyroid hormones. Therefore, the structural characteristics of the lysosomal system are described and discussed in terms of their relationship to the function of these adenomas.

## Material and Methods

12 of 71 human thyroid adenomas included in this series were composed of cells which, based on lightmicroscopic, immunohistochemical and electron microscopic findings represented a specific cytological differentiation.

Scintigraphically all tumors were inactive on scan. For details of tissue preparation see Dralle and Böcker (1977).

## Results

Specific differentiation was seen in the following types: 2 oxyphilic adenomas, 2 clear cell adenomas, 5 adenomas consisting of mitochondrion-rich cells and 3 consisting of ergastoplasm-rich cells. The tumor architecture varied considerably within each adenoma, nevertheless, the cytological features of each were sufficiently different to justify a distinction.

### *Light Microscopic Observations*

In the two instances in which *clear cell adenomas* were examined microscopically the tumor cells were arranged in compact cords (Fig. 1a). The hyperchromatic nuclei were small and round. There was an abundance of cytoplasm which contained small acidophilic granules. Small follicle lumina filled with colloid were seen occasionally.

Adenomas composed of *ergastoplasm-rich cells* showed a predominantly micro-to normofollicular architecture (Fig. 1b). The follicle lumina did not contain colloid. The cytoplasm of the tumor cells was somewhat less abundant and had an acidophilic granular appearance. Nuclei were small or medium sized with moderate amount of chromatin.

*Oxyphilic adenomas* (Fig. 1c) showed a predominantly trabecular pattern with the occasional formation of small follicles, the lumina of which contained only clumps of colloid. The polygonal cells were the largest in the whole series, with fine granular cytoplasm and distinct cell borders. A characteristic feature was a nuclei tendency to chromatin agglomeration with the formation of irregular configurations.

Adenomas of *mitochondrion-rich cells* (Fig. 1d) exhibited a microfollicular pattern of growth, the tumor cells had a cuboidal shape. The cytoplasm showed an intensively acidophilic granular staining especially at the cell base. The nuclei were relatively large and hyperchromatic.

*Immunohistochemical preparations* showed typical thyroglobulin content and distribution patterns in the four adenoma types.

Nearly all *clear cells* contained small amounts of thyroglobulin located within small cytoplasmic granules (Fig. 2). The small follicle lumina usually showed a moderately intense staining.

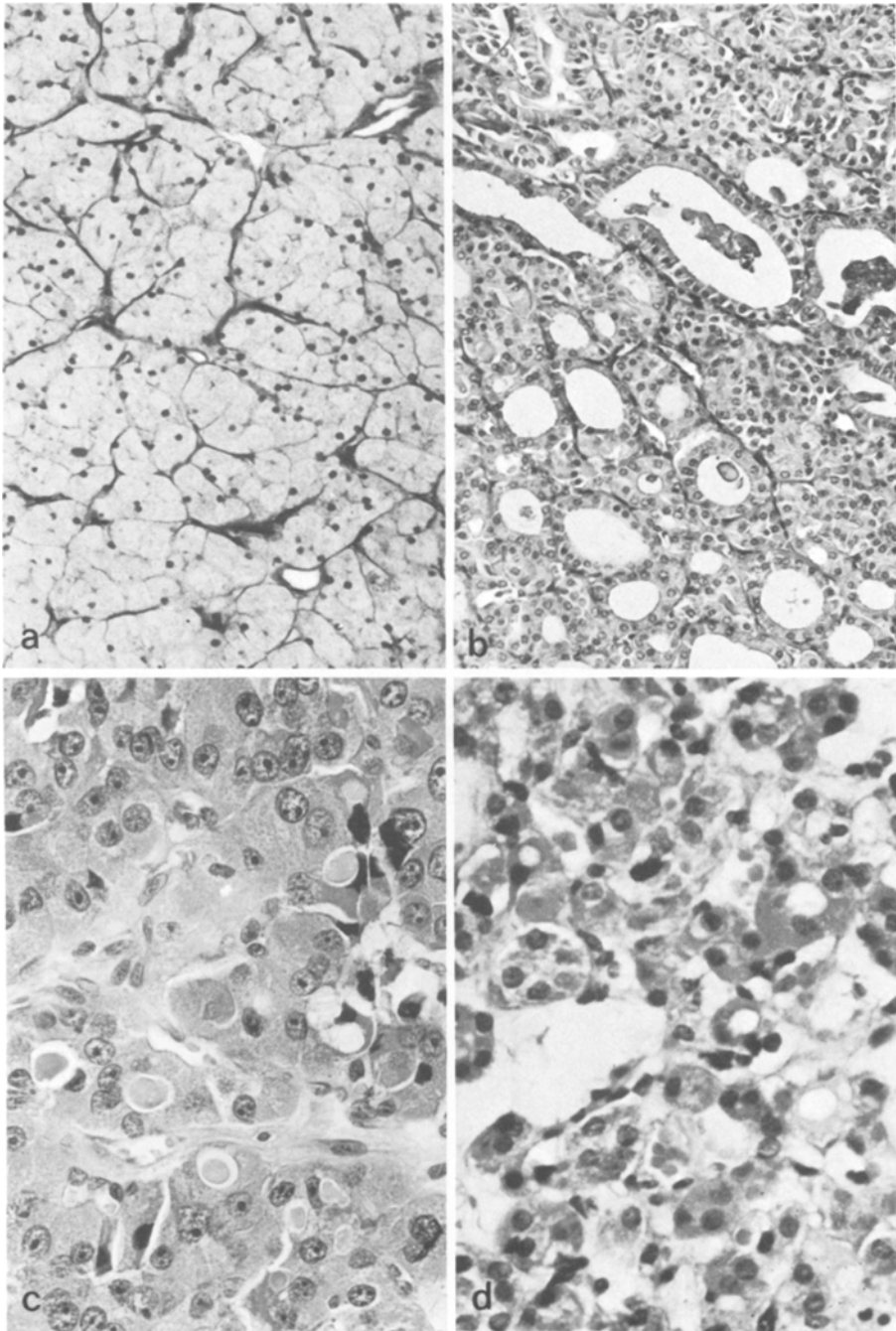
The *ergastoplasm-rich cell* was characterized by the extraordinary abundance of cytoplasmic and the lack of extracellular thyroglobulin with empty follicle lumina (Fig. 3).

The *oxyphilic adenomas* contained a few cells which showed large amounts of thyroglobulin (Fig. 4). However, in most tumor cells no thyroglobulin was detectable.

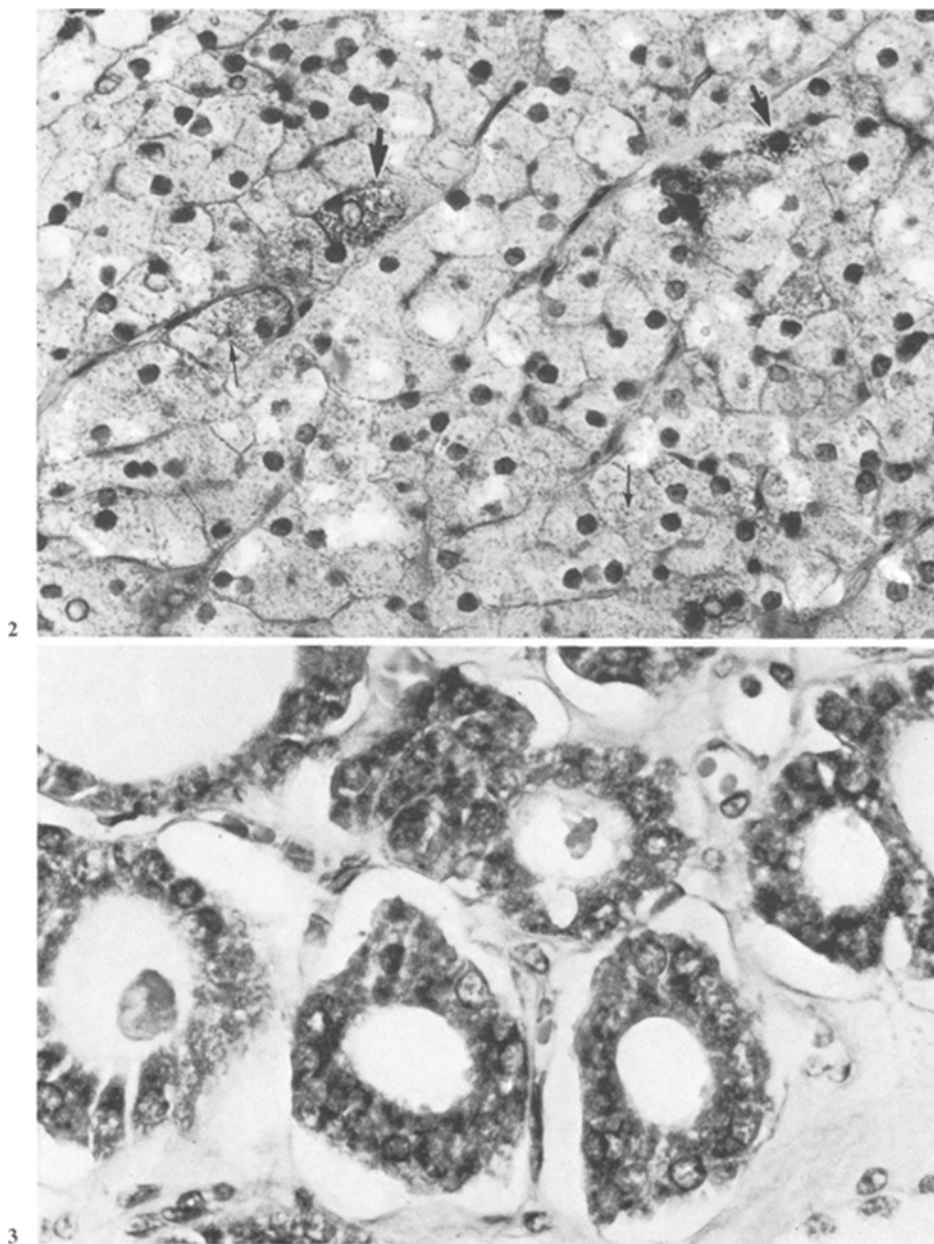
In *mitochondrion-rich cell adenomas* thyroglobulin was predominantly located in the apical portion of the cytoplasm, without sharp demarcation from the thyroglobulin of the follicle lumina (Fig. 5).

### *Electron Microscopic Observations*

The ultrastructure of the four cell types provided a rationale for the histological and the immunohistochemical observations. The fine structure of the *clear cell adenomas* has recently been described by Stoll and Lietz (1973). The cytoplasm contains many agranular empty vesicles of different sizes. Small areas of cytoplasmic matrix are occupied by oval or round cisternae of the rough endoplasmic reticulum (Fig. 6), mitochondria and extraordinary large lysosomes with a median diameter of 500 nm. These areas probably correspond to the immunohistochemically demonstrated thyroglobulin granules observed with the light microscope.

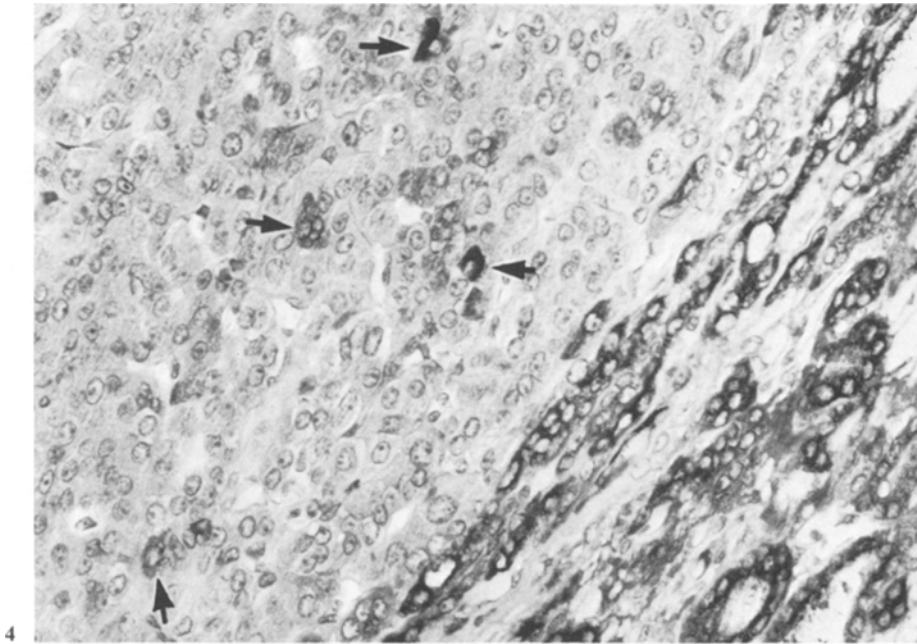


**Fig. 1a–d.** Histological aspects of adenomas with specific cytotypic differentiation. **a** Clear cell adenoma. **b** Ergastoplasm-rich cell adenoma. **c** Oxyphilic adenoma. **d** Mitochondrion-rich cell adenoma.  $\times 250$

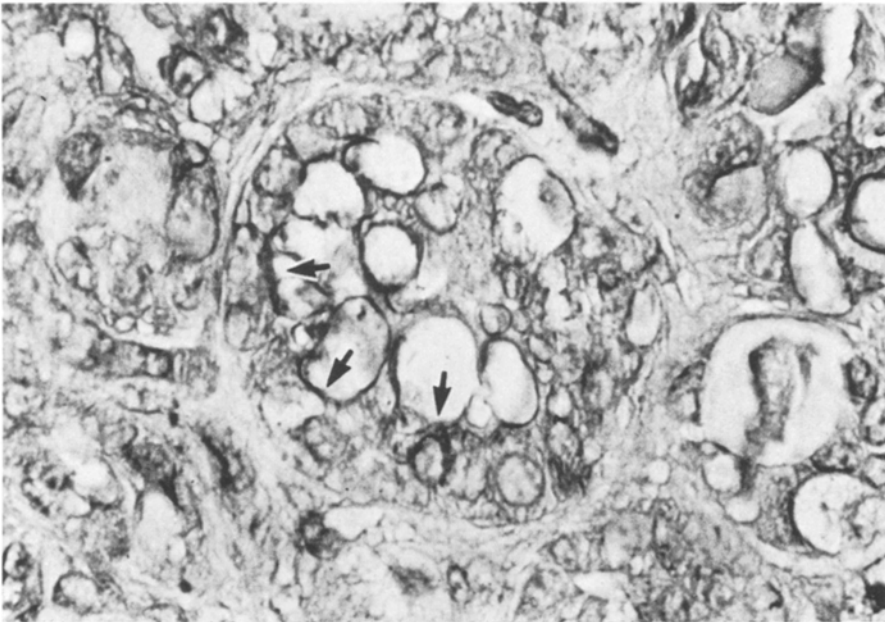


**Fig. 2.** Clear-cell adenoma. Immunohistochemical demonstration of thyroglobulin. Tumor cells reveal thyroglobulin in very small granules (*arrows*).  $\times 250$

**Fig. 3.** Ergastoplasm-rich-cell adenoma. Immunohistochemically the tumor shows a large amount of thyroglobulin which is almost completely confined to tumor cells.  $\times 400$



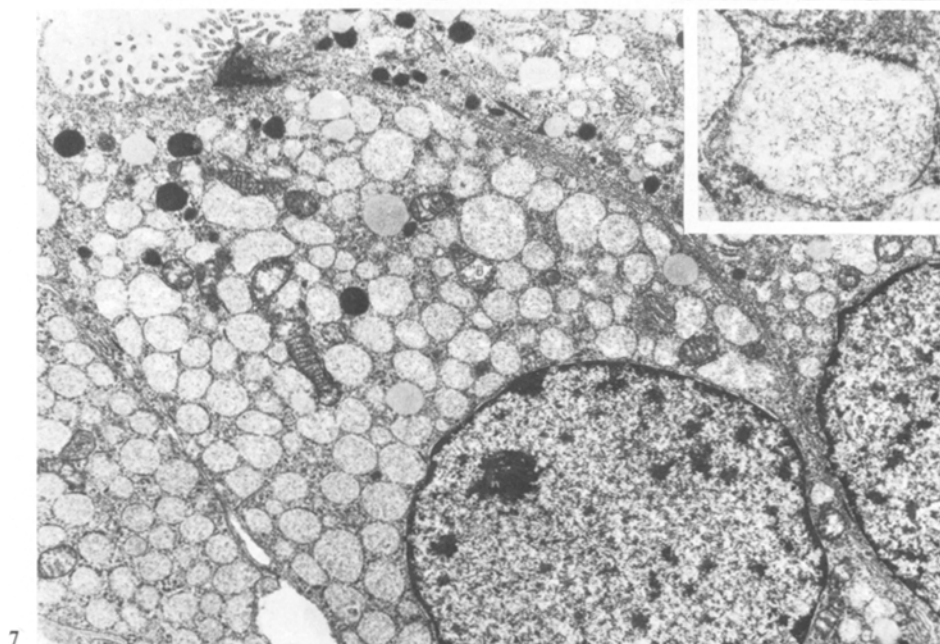
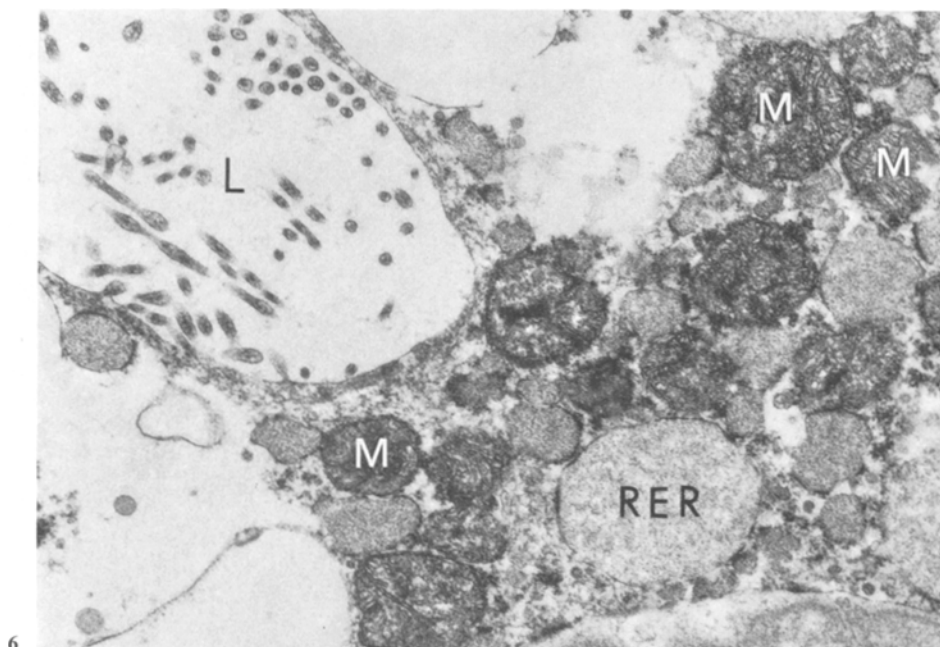
4



5

**Fig. 4.** Oxyphilic adenoma. Only few cells within the tumor contain thyroglobulin (*arrows*). On the right side normal, slightly compressed thyroid tissue is seen, the follicle cells of which contain a large amount of thyroglobulin. PAP-method.  $\times 250$

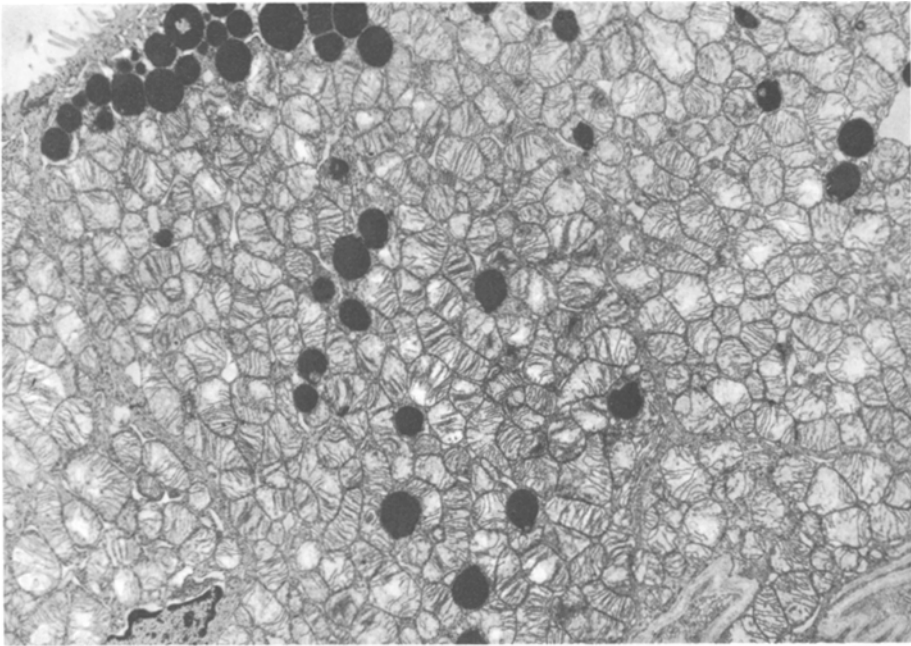
**Fig. 5.** Mitochondrion-rich cell adenoma. Immunohistochemically a moderate amount of thyroglobulin (*arrows*) can be localized within tumor cells and in the follicle lumen.  $\times 540$



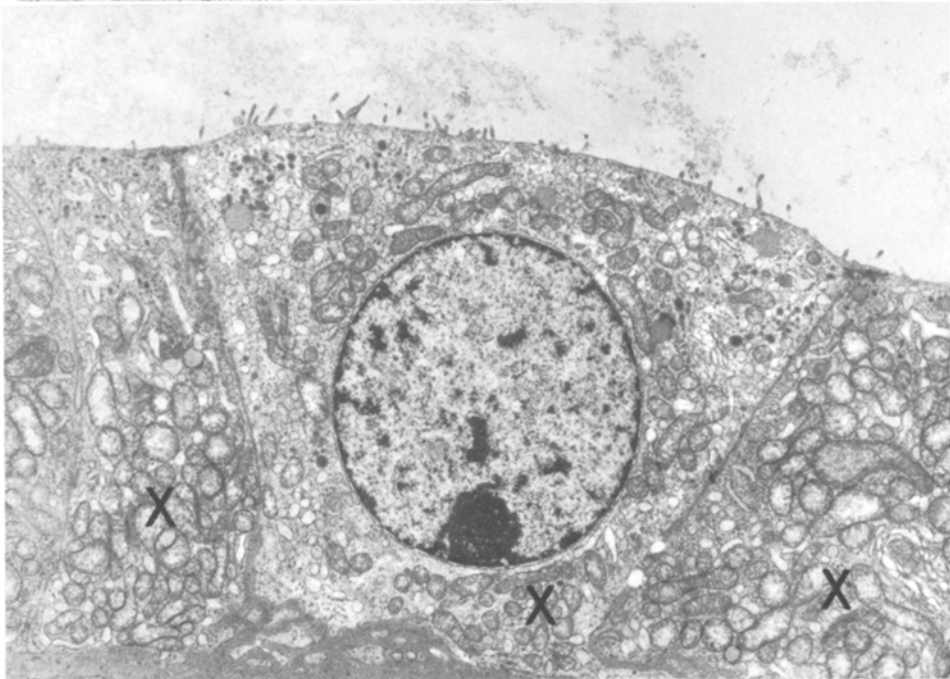
**Fig. 6.** Clear-cell adenoma. Apical portion of tumor cell with dilated cisternae of rough endoplasmic reticulum (*RER*) and mitochondria (*M*). *L* follicle lumen.  $\times 21,000$

**Fig. 7.** Ergastoplasm-rich cell adenoma. Tumor cell with numerous round cisternae of rough endoplasmic reticulum and large apically located lysosomes.  $\times 7500$ . *Inset*: higher magnification of a cisterna.  $\times 23,000$





8

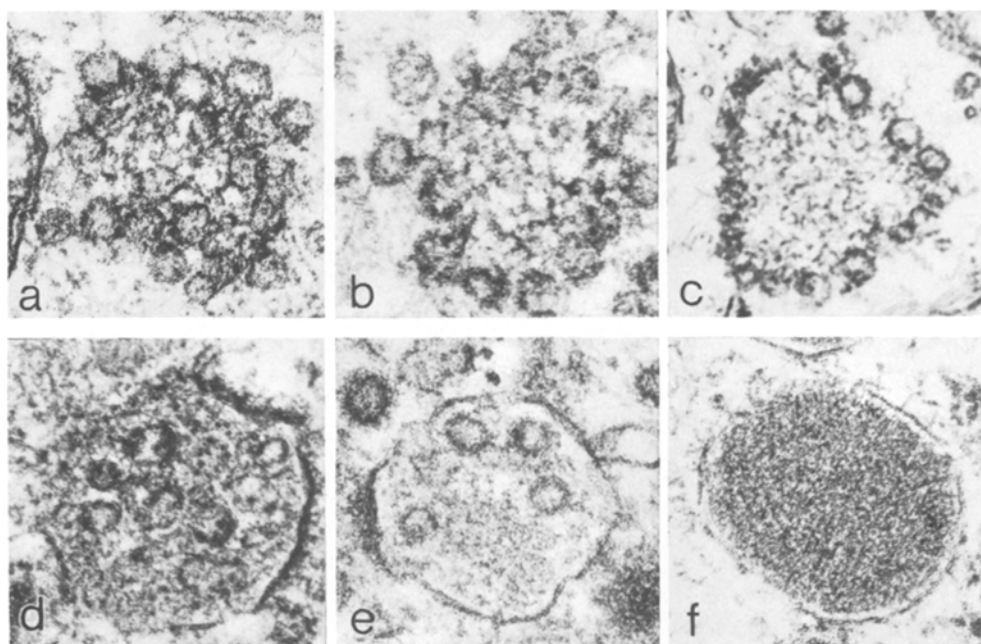


9

**Fig. 8.** Oxyphil-cell adenoma. Tumor cell with numerous mitochondria and large lysosomes. No other cell organelles.  $\times 5900$

**Fig. 9.** Mitochondrion-rich cell adenoma. In contrary to oxyphilic adenoma cells, these tumor cells contain only small lysosomes and many cytoplasmic vesicles. Rough endoplasmic reticulum is well developed. In many tumor cells the mitochondria are basally located. (X).  $\times 5800$



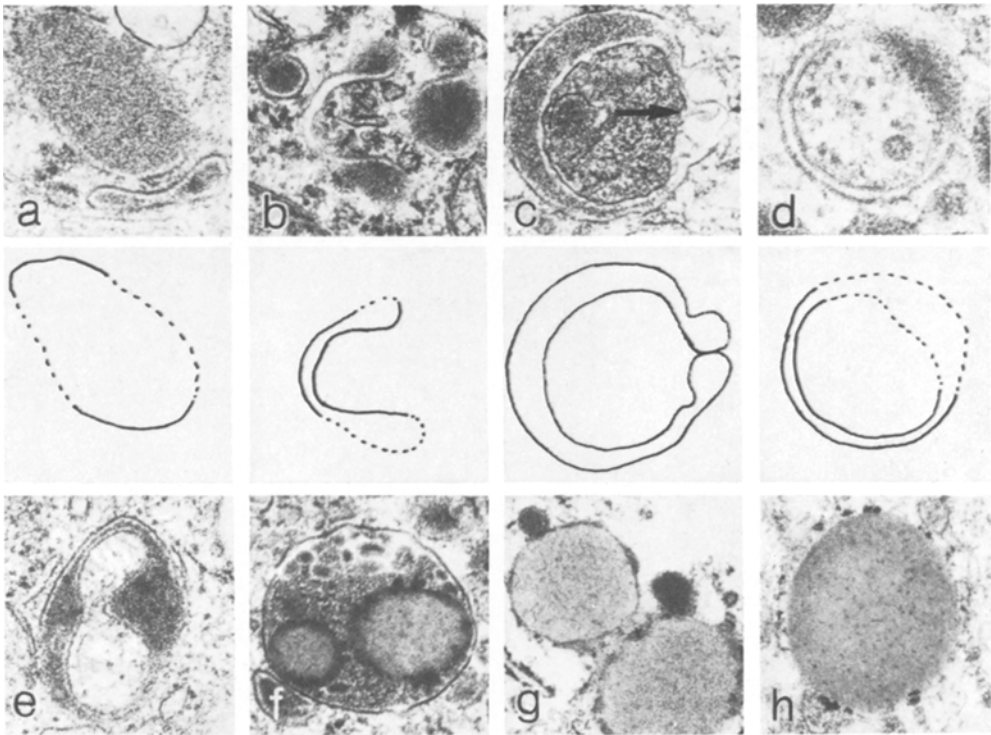


**Fig. 10a-f.** Stages in the formation of lysosomes from small Golgi vesicles. **a** Small cytoplasmic vesicles. **b** and **c** The membranes seem to dissolve and form a new common membrane (**d**). By inclusion of small vesicles, a multivesicular body results (**e**). **f** Lysosome.  $\times 75,000$

The ergastoplasm-rich cell (Fig. 7) contains cytoplasm full of ergastoplasmic cisternae with a finely granular colloid-like material. Golgi apparatus and cytoplasmic vesicles are barely developed. Lysosomes, which occur in moderate number in the apical cytoplasm, are large and correspond in size to the lysosomes of oxyphil and clear cells.

The ultrastructural features of *oncocytes* (Fig. 8) are well known from previous publications (Hamperl, 1950, 1962; Heimann et al., 1973; Valenta et al., 1974). Mitochondria are present in large number. The large lysosomes, mainly located apically, have a median diameter of 370 nm. All other cell organelles are present in small numbers.

Harcourt-Webster (1968) and Tremblay and Pearse (1960) in a histochemical study were able to distinguish a particular cell from the Hürthle-cell. It was characterized by an abundance of mitochondria and by being smaller than oxyphil cells and was called the *mitochondrion-rich cell* (Fig. 9). The present immunohistochemical and electron microscopical analysis of adenomas confirms the existence of this cell type. When compared with the typical oxyphil cell the mitochondrion-rich cell is characterized by a smaller number of usually basally and laterally localized mitochondria, and the presence of many small lysosomes with a diameter of 160 nm and few lamellae of the rough endoplasmic reticulum in the cytoplasm. In contrast to other adenomata, numerous small cytoplasmic vesicles are seen throughout the cytoplasm.



**Fig. 11.** Cellular autophagy. Suggested steps in the formation of autophagic vacuoles. **a** normal lysosome. **b** and **c** increasing invagination of the lysosomal membrane with segregation of a cytoplasmic area. **d** and **e** By fusion of the touching ends (arrows) an autophagic vacuole results. **f** Transformation to secondary lysosomes by dissolution of the inner membrane. **f, g, h** Lipofuscin granules.  $\times 49,000$

### *The Lysosomal System*

In the thyroid gland the lysosomal enzymes play a part in the elaboration of hormones (Wollman et al., 1964; Seljelid, 1975) and also with autophagic processes (Nunez and Becker, 1970). From their general morphological aspects, it is suggested that the oxyphilic, the clear cell and the ergastoplasm-rich cell adenomas are functionally inactive: for this reason these cells seem suitable for a study of those aspects of the lysosomal system that are not concerned with the enzymatic digestion of thyroglobulin.

According to our ultrastructural and cytochemical studies the lytic system of these cells can be divided into the following components (Fig. 10): 1. Small cytoplasmic vesicles with contents of low electron density, which probably originate from cisternae of the Golgi field. 2. Typical lysosomal dense bodies of varying size. 3. Multivesicular bodies and 4. autophagic vacuoles. Clusters of small smooth surfaced vesicles are frequently seen at the ends of the Golgi cisternae. Some have contents of high electron density and seem to represent small primary lysosomes with diameters of 60–110 nm. Another type of small

vesicle with contents of low electron density is also observed. From these vesicles, multivesicular bodies seem to evolve in several steps (a) small vesicles, which probably originate from Golgi cisternae, are first localized in small groups. (b) The membranes of the outer vesicles apparently fuse and form a large vesicle, the newly formed membrane of which has irregular outlines with patchy thickening. As this larger body usually includes a few of the original small vesicles, a multivesicular body appears. Acid phosphatase preparations clearly reveal a number of small vesicles within the multivesicular bodies. These show a positive staining reaction, indicating that they belong to the lytic system of the cells.

*Autophagosomes and/or autophagolysosomes* are rarely encountered in adenomas. The membrane kinetics of autophagic vacuoles are unknown (Ericsson, 1969; Pfeifer, 1976). During the present study images were found (Fig. 11) which resemble those found by Nunez and Becker (1970) in the bat thyroid and which suggest that autophagic vacuole seems to result from an invagination of the lysosomal membrane itself, with lysosomal enzymes, located between the two membranes, forming the autophagic vacuole. Transformation to the autophagolysosomes seems to result from dissolution of the inner membrane.

## Discussion

In our analysis of 71 human thyroid adenomas the following five cell types were distinguished:

1) Main (principle) cell (including "undifferentiated" cells in trabecular adenomas, atrophic cells in macrofollicular and hypertrophic cells in autonomous adenomas).

2) Oxyphil (oncocytic) cell = Askanazy cell (Fig. 12a).

3) Clear cell (Fig. 12b).

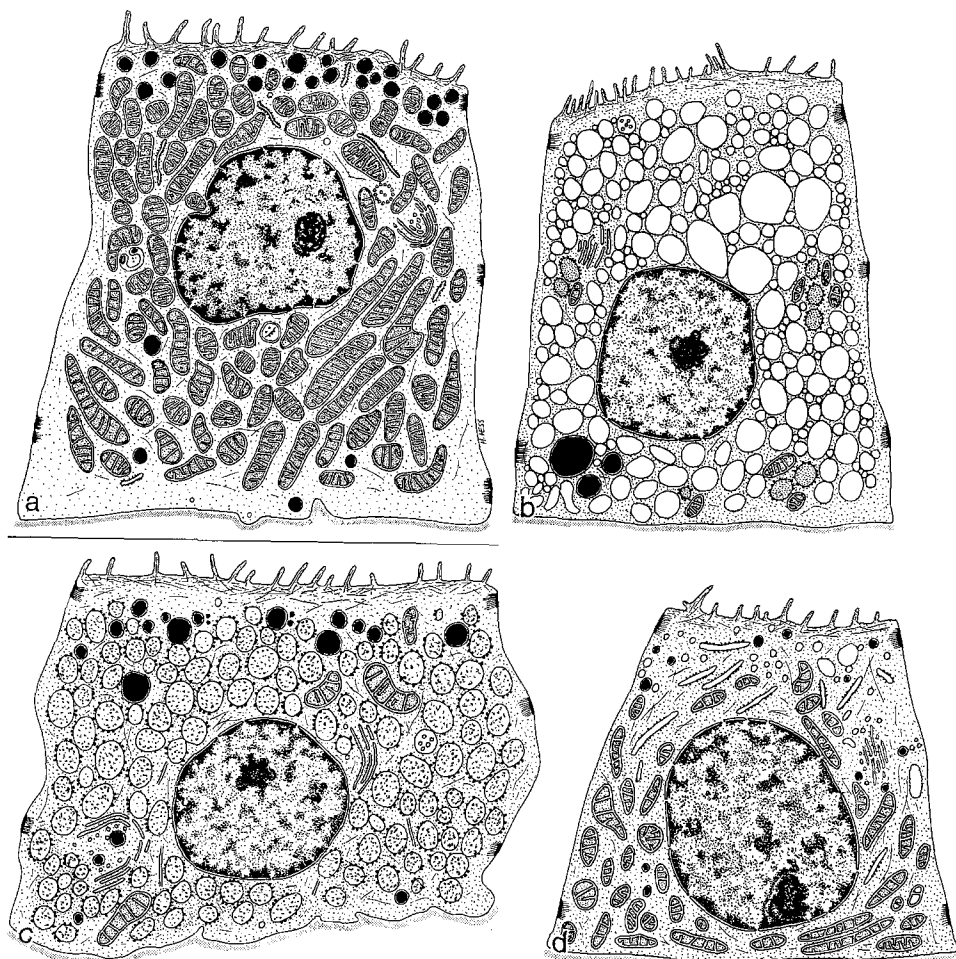
4) Ergastoplasm-rich cell (Fig. 12c).

5) Mitochondrion-rich cell (Fig. 12d).

87% of the tumors studied were main cell adenomas, as already described (Dralle and Böcker, 1977). 17% of the adenomas (cell types 2–5) analysed in this study revealed architectural and cytologic features which were easily distinguished by light microscopic, immunohistochemical and by electron microscopic features. These were regarded as adenomas with specific cytological differentiation.

Cell types 2–5 were characterized by a selective increase of *one* cell organelle with a decrease in number and/or in size of all or most other cell organelles. The organelles involved were the mitochondria in the oxyphil cell and mitochondrion-rich cell, the smooth endoplasmic reticulum in the clear cell and the rough endoplasmic reticulum in the ergastoplasm-rich cell.

The best studied example is the *oxyphil cell*. This cell type can easily be recognized light-microscopically by its large size, eosinophilic granular cytoplasm (see recent review by Roediger, 1975), its high content of oxydative mitochondrial enzymes, basophilic granularity of the cytoplasm after phosphotungstic-acid-hematoxyline staining, and by reduction and/or lack of thyroglobulin synthesis.



**Fig. 12a-d.** Schematic diagram of the different cell types. **a** Oxyphil cell, **b** clear cell, **c** ergastoplasm-rich cell, **d** mitochondrion-rich cell

Our immun-histochemical findings clearly indicate that most of the cells of oxyphilic adenomas do not produce any thyroglobulin. But there are few cells which showed a very intensive reaction and which were considered to be main cells. The nature of the process leading to oxyphil cell change is unknown. Recently it has been suggested that the oxyphil cell represents a metaplastic follicle cell, in contrast to the earlier views of Hamperl (1950).

In an ultrastructural analysis Stoll and Lietz (1973) have analyzed the *clear cell*. It is characterized by an abundance of smooth surfaced vacuoles of varying size which either develop from the Golgi field or from endocytotic vesicles. Whereas earlier authors (Langhans, 1907) have suggested an origin of the clear cell from the parathyroid, most authors now (Hedinger et al., 1967) accept that this cell is derived from the follicle cell. The present study revealed cytologic

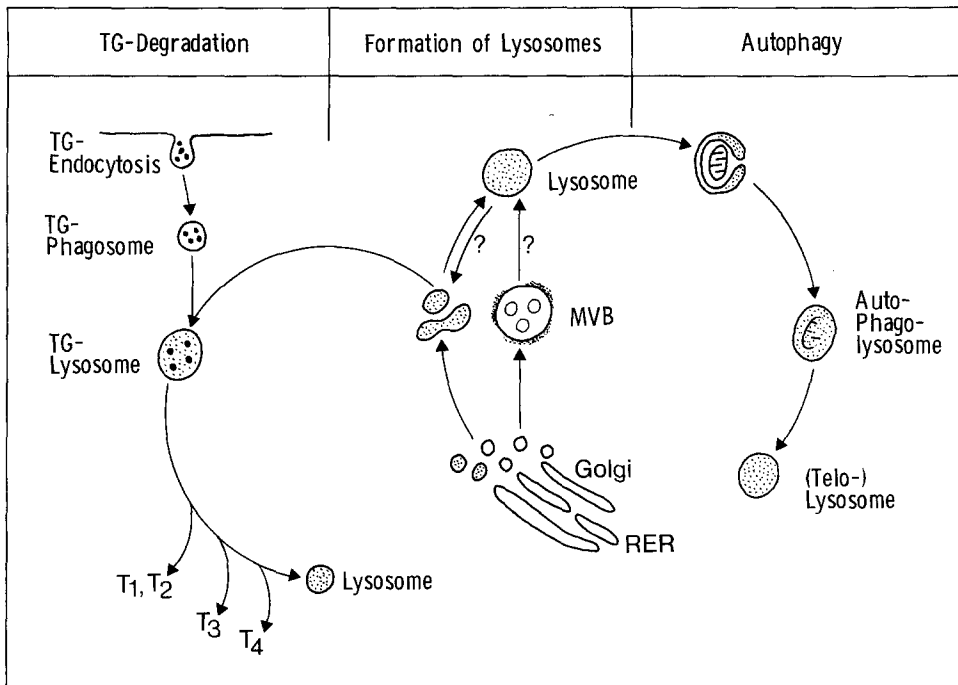
features supporting the latter hypothesis. These features are the immunohistochemically demonstrated thyroglobulin and the electronmicroscopically observed profiles of rough endoplasmic reticulum, Golgi complexes and lysosomes in most of the tumor cells.

The *ergastoplasm-rich cell* has not been previously described. Morphologically the cell is characterized by its abundance of cisternae of the rough endoplasmic reticulum which are arranged in round profiles. They contain a large amount of thyroglobulin, which can be demonstrated immunohistochemically. Lightmicroscopically the cell body is smaller than the oxyphil cell, the cytoplasm is eosinophilic and granular so that our original histological diagnosis was oxyphilic adenoma. On PAS-staining these tumors show a slightly positive cytoplasmic reaction but even in areas with well formed follicles there is characteristically no or very little colloid. On the basis of the morphological features, it is suggested that this cell type, like the oxyphil and clear cell, represents a well defined and easily recognized metaplastic follicle cell.

In contrast to the cells described above, the *mitochondrion-rich cell* remains somewhat enigmatic in its functional activity and cytogenesis. From immunohistochemical findings it is evident that this cell synthesizes and secretes thyroglobulin into the follicle lumen. In addition, it is characterized by its large number of mitochondria, its many small lysosomes (in contrast to oxyphil cells), some stacks of rough endoplasmic reticulum and a moderately prominent Golgi field. It is our contention that this cell type corresponds to the mitochondrion-rich cell which according to the histochemical studies (DPN-diaphorase, succinate-dehydrogenase, TPN-diaphorase, TPN-linked isocitrate dehydrogenase activity) of Tremblay and Pearse (1960), Harcourt-Webster and Scott (1966) and Valenta et al. (1974) is found in a variety of thyroid disorders. To the best of our knowledge it is the first time that this cell has been found to be the predominant cell type in a human thyroid adenoma.

The morphological findings obtained suggest that cell types 2-4 are completely inactive hormonally and, furthermore that they do not even degrade thyroglobulin. Direct evidence supporting hormonal inactivity is provided by lack of thyroglobulin in oxyphilic adenoma cells and decrease in the number of those organelles involved in thyroglobulin-transport, i. e. the Golgi complex and the small cytoplasmic vesicles (exocytic and endocytic). An interesting finding is that in cell types 2-4 the lysosomes when compared with hormonally active cells (autonomous adenomas and thyroids in thyrotoxicosis) are very large (360 nm in diameter compared with 250 nm in hormonally active cells).

As the oxyphil cell is generally a non-thyroglobulin synthesizing and degrading cell, and as the lysosomes in this cell are very numerous, it may be hypothesized that large lysosomes represent the morphological equivalent of inactivity and possibly represent "physiological" enzyme storage. In the normal cell this is transformed to smaller lysosomes if required. It is suggested that the lysosomes are derived from the Golgi cisternae (Sobel, 1962; Seljelid, 1967; Coleman et al., 1968) but another mode of formation of the typical thyroid lysosomes was suggested by Nunez and Becker (1970). These authors are of the opinion that they develop from the multivesicular body, which in turn derives from the Golgi field (see Fig. 13). From our own electron microscopic findings it



**Fig. 13.** Schematic diagram of the lysosomal system with thyroglobulin-degradation, formation of lysosomes and cellular autophagia

is suggested that both pathways are possible in thyroid adenomas. Some of the images obtained seem to suggest that the outer membrane of the multivesicular body develops from the small vesicles by a very complex process which is not clearly understood and deserves further study. Nevertheless, histochemical studies (Böcker, 1978) clearly indicate that multivesicular bodies belong to the lytic system of the adenoma cell.

The mode of formation of *autophagic vacuoles* has been seen in a number of different cell types (ref. see Pfeifer, 1976). Most hypothesis assume that the membranes surrounding autophagolysosomes developed from the endoplasmic reticulum (Hugon and Borger, 1965; Glinsman and Ericsson, 1966; Cole et al., 1971; Ericsson et al., 1975) and/or from Golgi membranes (de Duve and Wattaux, 1966), lysosomal membranes (Novikoff et al., 1964; Seljelid, 1966; Seljelid and Ericsson, 1966; Bartok et al., 1967; Dixon, 1967) or de novo synthesis (Ashford and Porter, 1962; Napolitano, 1963). Nunez and Becker (1970) in their study of the bat thyroid suggest that autophagic vacuoles may also develop from the lysosomes themselves. From our own observations the same mechanism seems to occur in human thyroid adenomas.

*Acknowledgments.* The authors wish to thank Mrs. M. Fischer and Mrs. U. Zeiger for their excellent technical assistance and Mr. H. Hess for drawing the schematic diagrams.

## References

- Ashford, T.P., Porter, K.R.: Cytoplasmic components in hepatic cell lysosomes. *J. Cell Biol.* **12**, 198–202 (1962)
- Askanazy, M.: Pathologisch-anatomische Beiträge zur Kenntnis des Morbus Basedowii insbesondere über die dabei auftretende Muskelerkrankung. *Dtsch. Arch. klin. Med.* **61**, 118–186 (1898)
- Bartok, I.V., Totovic, V., Gedigk, P.: Über die Entstehung der peribiliären dichten Körper der Leberzellen. Untersuchungen in der präregeneratorischen Phase nach subtotaler Hepatektomie. *Virchows Arch. Path. Anat.* **343**, 1–19 (1967)
- Cole, S., Matter, A., Karnowsky, M.J.: Autophagic vacuoles in experimental atrophy. *Exp. Molec. Pathol.* **14**, 158–175 (1971)
- Coleman, R., Evenett P.J., Dodd, J.M.: Ultrastructural observations on some membranous cytoplasmic inclusion bodies in follicular cells of experimentally-induced goitres in tadpoles and toads of *Xenopus laevis*. *Duadin. Z. Zellforsch.* **84**, 497–505 (1968)
- De Duve, Ch., Wattaux, R.: Function of lysosomes. *Ann. Rev. Physiol.* **28**, 435–492 (1966)
- Dixon, J.S.: Phagocytic lysosomes in chromatolytic neurones. *Nature* **215**, 657–658 (1967)
- Dralle, H., Böcker, W.: Immunohistochemical and electron microscope analysis of adenomas of the thyroid gland. I.A. comparative investigation of hot and cold nodules. *Virchows Arch. A Path. Anat. and Histol.* **374**, 285–301 (1977)
- Ericsson, J.L.E., Trump, B.F., Weibel, J.: Electron microscopic studies of the proximal tubule of the rat kidney. II. Cytosegrosomes and cytosomes: their relationship to each other and to the lysosome concept. *Lab. Invest.* **14**, 1341–1365 (1965)
- Ericsson, J.L.E.: Mechanism of cellular autophagy. In: *Lysosomes in biology and pathology II*, pp. 345–387 (Dingle, J.T., Fell, H.B., eds.). Amsterdam, London: North Holland Co., 1969
- Glinksman, W.H., Ericsson, J.L.E.: Observations on the cellular organization of hepatic parenchymal cells. II. Evolution of reversible alteration induced by hypoxia. *Lab. Invest.* **15**, 762–777 (1966)
- Hamperl, H.: Oncocytes and the so-called Hürthle-cell tumor. *Arch. Path.* **49**, 563–567 (1950)
- Hamperl, H.: Onkozyten und Onkozytome. *Virchow Arch. Path. Anat.* **335**, 452–483 (1962)
- Harcourt-Webster, J.N.: The role of enzyme histochemistry as a tool in the study of thyroid neoplasms. In: *Thyroid neoplasia*, pp. 443–457 (Young, S., Imman, D.R., eds.). London, New York: Academic Press, 1968
- Hedinger, Ch., Corbat, F., Egloff, B.: Schilddrüsenmetastasen hypernephroider Nierencarcinome. *Schweiz. Med. Wochenschr.* **97**, 1420–1426 (1967)
- Heimann, P., Ljunggren, J.G., Löwhagen, T., Hjern, B.: Oxyphilic adenoma of the human thyroid gland. *Cancer* **31**, 246–254 (1973)
- Hugon, J., Borgers, M.: Etude morphologique et cytochimique des catolysosomes de la crypte duodénale. *J. Microscopie* **4**, 608–618 (1968)
- Kennedy, J.S., Thompson, J.A.: The changes of the thyroid after irradiation with <sup>131</sup>I or partial thyroidectomy for thyrotoxicosis. *J. Pathol.* **112**, 65–82 (1974)
- Klinck, J.H., Örtel, J.E., Winship, T.: Ultrastructure of the normal human thyroid. *Lab. Invest.* **22**, 2–22 (1970)
- Langhans, T.: Über die epithelialen Formen der malignen Struma. *Virchows Arch. Path. Anat.* **189**, 69–189 (1907)
- Lennox, B.: The large cell small acinar thyroid tumor of Langhans and the incidence of related cell groups in the human thyroid. *Path. Bacteriol.* **60**, 295–305 (1948)
- Michel-Bechet, M., Valenta, L.J., Kymel, F.: Further advances in thyroid research. Wien: Wien. Med. Akad. 1971
- Napolitano, L.: Cytolysosomes in metabolically active cells. *J. Cell Biol.* **18**, 478–481 (1963)
- Novikoff, A.B., Essner, E., Quintana, N.: Golgi apparatus and lysosomes. *Federation Proc.* **23**, 1010–1022 (1964)
- Nunez, J., Becker, D.V.: Secretory processes in follicular cells of the bat thyroid. *Am. J. Anat.* **129**, 369–398 (1970)
- Pfeifer, U.: Lysosomen und Autophagie. *Verh. Dtsch. Ges. Path.* **60**, 28–64 (1976)
- Roediger, W.E.M.: The oxyphil and the C cell of the human thyroid gland. *Cancer* **36**, 1758–1770 (1975)
- Seljelid, R.: On the origin of colloid droplets in thyroid follicle cells. *Exper. Cell Res.* **41**, 688–691 (1966)



- Seljelid, R., Ericsson, J.L.E. In: Electron microscopy (Uyeda, R., ed.) Proceedings of the 6<sup>th</sup> International Congress for Electron Microscopy, Kyoto, Japan, 1966. Citation in: Ericsson, J.L.E., Trump, B.F., Weibel, J.: Electron microscopic studies of the proximal tubule of the rat kidney. II. Cytosegrosomes and cytosomes: their relationship to each other and to the lysosome concept. *Lab. Invest.* **14**, 1341–1365 (1965)
- Seljelid, R.: Endocytosis in thyroid follicle cells. I. Structure and significance of different types of single membrane limited vacuoles and bodies. *J. Ultrastruct. Res.* **17**, 195–219 (1967)
- Seljelid, R.: Thyroid lysosomes in health and disease. In: Pathobiology of cell membranes, Trump, B.F., Arstila, A.U. eds. pp. 325–381. New York, San Francisco, London: Academic Press, 1975
- Sobel, H.J.: Relationship of three lysosomal enzymes to the Golgi zone and secretory activity in the rat pituitary and thyroid glands. *Anat. Rec.* **143**, 389–393 (1962)
- Stoll, W., Lietz, H.: Zur Kenntnis und Problematik des hellzelligen Adenomas in der Schilddrüse. *Virchows Arch. Abt. A Path. Anat.* **361**, 163–173 (1973)
- Tonietti, G., Bachieri, L., Salabe, G.: Papillary and microfollicular carcinoma of the human thyroid. *Arch. Path.* **84**, 601–614 (1967)
- Tremblay, G., Pearce, A.G.E.: Histochemistry of oxidative enzyme systems in the human thyroid with special reference to Askanazy cells. *J. Pathol. Bacteriol.* **80**, 353–358 (1960)
- Valenta, L., Michel-Bechet, M., Warshaw, J.B., Maloof, F.: Human thyroid tumors composed of mitochondrion-rich cells: electron microscopic and biochemical findings. *J. Clin. Endocrinol.* **39**, 719–733 (1974)
- Wegelin, C.: Die Parastruma maligna (Langhans). In: Handbuch der speziellen pathologischen Anatomie und Histologie (Henke and Lubarsch, eds.), Vol. VIII, pp. 1–547, Berlin: Springer 1926
- Wollman, S.H., Spicer, S., Burnstone, M.: Localization of esterase and acid phosphatase in granules and colloid in rat thyroid epithelium. *J. Cell. Biol.* **21**, 191–201 (1964)

Received May 26, 1978